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Research of the mechanism of acetic acid fluorescence quenching^{*}

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Abstract: Analyses of the mechanisms of acetic acid emitting fluorescence and fluorescence quenching have been made in theory. The results indicate that acetic acid solutions can emit visible fluorescence when induced by 253.7nm UV light, and fluorescence quenching occurs along with the alteration of solution concentration. It is thought that fluorescence dynamic quenching process could take into account the intersystem transition of (n, π^*) to (π, π^*), and the quenching constant is calculated in this paper. Investigation on the native fluorescence spectrum and its characteristics will contribute to the study of other molecules fluorescence spectra when acetic acid serves as solute, hydrolysis catalyst and food additive.

Key words: acetic acid; fluorescence spectrum; fluorescent quenching; ultraviolet light-induced

乙酸溶液荧光猝灭的机理研究

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摘要: 从理论上对紫外光激励乙酸产生荧光及荧光猝灭机理分别进行了分析研究。结果表明, 乙酸溶液在 253.7nm 紫外光照射下可以发出明显的荧光, 而且随着溶液浓度的变化产生了荧光猝灭的现象。分析认为, 荧光猝灭的主要原因是动态猝灭过程——由最低激发单重态(n, π^*)转入三重态(π, π^*)所致, 并对猝灭常数进行了计算。研究乙酸本体荧光特性及荧光猝灭机理可为其作为猝灭剂、溶剂、催化剂和食品添加剂时其它分子荧光光谱的研究提供参考。

关键词: 乙酸; 荧光光谱; 荧光猝灭; 紫外光激励

中图分类号: Q632 **文献标识码:** A

Instruction

Acetic acid has been extensively studied as an organic solvent, where it is mostly used as a hydrolysis catalyst, fluorescence quenching, and food additive, and the characters of other molecules fluorescence have been well studied when acetic acid serves as an additive. It is well known that acetic acid plays an important role in colposcopic diagnostics in order to detect some cervical abnormalities, making them white and more noticeable by the human eye. Zelenchuk studied about fluorescence spectra

of freshly excised cervical tissue and the data in this study from non-neoplastic cervical tissue reveal changes after the application of acetic acid^[1]. Harris research demonstrated that in the case of acetic acid fixation sperm visualization became easier and could make the difference in fluorescence coloration between sperm and cell cytoplasm^[2]. Acetic acid also has been well studied when it serving as food additive. Acetic acid is a potential inhibitor of yeast growth and survival^[3]. Its inhibition mechanism was generally assumed to involve the passive diffusion of acetic acid in its undissociated form across the plasma membrane of the cell. And the reason of causing intracellular acidification was that the undissociated acetic acid dissociates because of the higher intracellular pH^[4,5]. When serving as fluorescence quenching, acetic acid quenching mechanism has been studied. Grousi's found that formaldehyde

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could emit a strongly fluorescence in investigating fluorometric determination of formaldehyde and fluorescence was quenched by acetic acid. Quintero *et al.* pointed out that fluorescence dynamic quenching process could represent a serious inconvenience in analytical fluorimetric techniques taking into account acetic acid being commonly used as a buffer and additional fluorescence quenching by H^+ ions could be involved in acid aqueous mediums^[6]. But in all of these studies, acetic acid was considered as a nonfluorescence emission matter.

Despite an extensive research on acetic acid when serving as solvent, we have never found any publication describing acetic acid native fluorescence emission. The main reason is that acetic acid doesn't absorb visible light, that is to say, it is optic transparent within this wavelength, so it has no effect on other molecules emitting fluorescence. However, acetic acid can absorb ultraviolet light; few researchers study whether acetic acid can emit fluorescence after absorbing these UV lights. Almost in all researches when acetic acid served as solvent, it is widely noted that fluorescence quenching is caused by acetic acid with dynamic and static processes. Therefore, it is very urgent to clarify the fluorescent character and the mechanisms of acetic acid fluorescence quenching. In this paper, fluorescence spectra of acetic acid are investigated and the dependence of fluorescence intensity and quenching phenomenon are presented.

1 Materials and experimental results

The experimental apparatus used to obtain the fluorescence and absorption spectra from different acetic acid aqueous solutions have been detailed in our earlier paper^[7]. All the reagents are with high purity. In the spectrofluorimetric measurements triply distilled water is used to dilute the sample to form different concentration solutions. In order to test fluorescence spectra character of acetic acid solutions with different concentrations, the sample is diluted to form from several thousandths till to several millionths. Then test the fluorescence intensity alteration and the degree of fluorescence quenching effect respectively.

According to our early research, the fluorescence intensity changes as the sample concentrations alter, fluorescence quenching occurs in some solutions and the fluo-

rescence intensity is so sensitive to the acetic acid concentration that fluorescent quenching occurs now and then.

2 Discussions

According to the experimental results given above we can conclude that acetic acid is able to radiate fluorescence when induced by 253.7nm ultraviolet light. When the exciting light transmits through the acetic acid solution, the energy of exciting light mostly converts fluorescence photon energy. Acetic acid molecule enables its primary fluorophore to absorb the energy of exciting light and then transfer it into fluorescence photon energy. Based on the principle of fluorescence emission, acetic acid molecule has fluorophore. In analytical chemistry, chromophore group is the undersaturated part of a molecule, which can absorb UV-VIS light and bring electron transitions. The chromophore in acetic acid molecule is $C=O$ ^[8] which has several types of electron transition such as $\sigma \rightarrow \sigma^*$, $n \rightarrow \sigma^*$ and $n \rightarrow \pi^*$ (See Fig. 1).

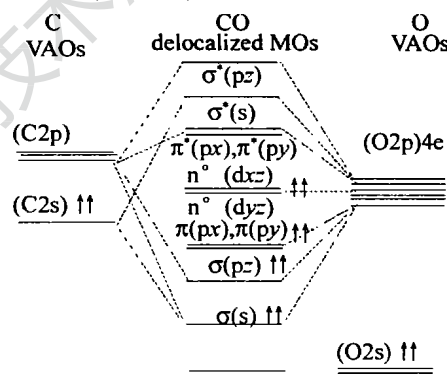


Fig. 1 Delocalized MO energy level diagram for C=O

Among these energy level transitions, the $n \rightarrow \sigma^*$, which is from nonbonding electron namely n electron to antibonding orbital σ^* , must absorb the lowest photon energy. Because electron transitions of $\sigma \rightarrow \sigma^*$, $n \rightarrow \sigma^*$ and $n \rightarrow \pi^*$ must absorb exciting light with wavelength less than 200nm, we can conclude that the sample excited by 253.7nm UV light results in $n \rightarrow \sigma^*$ electronic transition in chromophore $C=O$, that is to say, the n electrons in ground state absorb the photons energy and jump to excited state of σ^* orbit. In order to return to the ground state, the unstable electrons in excited state, losing its excess energy usually in the form of heat, transit to lower rotational and vibrational levels of excited state, and then return to the ground state and radiate fluorescence. Therefore the wavelength of the emitted fluorescence is

longer than the exciting light. From the different lower levels of the excited state, the excited electrons return to the ground state and emit fluorescence photon with different energy, thus the broadening fluorescence spectrum peak around 395nm.

To understand the effect of the quenchers on the fluorescence of the acetic acid, the data are analyzed conventionally by the Stern-Volmer equation and dynamic quenching process^[9]:

$$I_0/I_q = 1 + k_{SV}[Q] \quad (1)$$

Where I_0 and I_q are the unquenched and quenched fluorescence emission intensities respectively; k_{SV} is the Stern-Volmer constants.

The first excited singlet state of carbonyl compound is (n, π^*) and in this case the quantum yield of $S_1 \rightarrow T_1$ intersystem transition can be increased. Because $n \rightarrow \pi^*$ transition is partially forbidden, the antitransition process of $\pi^* \rightarrow n$ is also partially forbidden. As the result, the life in the first excited singlet state (n, π^*) is longer than that in triplet state (π, π^*) and the probability of intersystem transition of singlet-triplet is increased. Therefore, the first excited singlet states of acetic acid are very easy to transform to triplet and thus fluorescent quenching is easy to occur. According to the hypothesis of constant states, the concentration of excited-state molecule $^1M^*$ keep unvaried after being illuminated by excited light, and fluorescent quenching process caused by singlet-triplet transition are presented as follows:



The concentration $^1M^*$ with no fluorescent quenching is denoted as $[^1M^*]^0$ when singlet-triplet transition doesn't occur:

$$[^1M^*]^0 = \frac{I_a}{k_f + k_{ic} + k_{isc}} = \frac{I_a}{k_f + k_i} = \frac{I_a}{k_f + k_{ic}} \quad (5)$$

Where I_a is absorption speed or molecule activation created-speed, k_f is fluorescent speed constant, k_{ic} and k_{isc} are the constant of inner transform and intersystem transition speed. In the case of fluorescent quenching the singlet-

triplet transition, $[^1M^*]$ is maximal and $[^1M^*]$ is:

$$\phi_f^0 = \frac{k_f[^1M^*]^0}{I_a} \quad \text{and} \quad \phi_f = \frac{k_f[^1M^*]}{I_a}$$

represent the fluorescent quantum yield with and without existence of fluorescent quenching respectively. Based on the above relations, they are described as the following equation:

$$\phi_f^0 = \frac{k_f}{k_f + k_{ic}} \quad (7)$$

$$\phi_f = \frac{k_f}{k_f + k_{ic} + k_{isc}} \quad (8)$$

The ratio of ϕ_f^0/ϕ_f is described as:

$$\frac{F_0}{F} = \frac{\phi_f^0}{\phi_f} = \frac{k + k_{ic} + k_{isc}}{k + k_{ic}} = 1 + \frac{k_{isc}}{k + k_{ic}} \quad (9)$$

It can be obtained from Eq. (6) that the quenching constant k_{SV} is:

$$k_{SV} = \frac{k_{isc}}{k + k_{ic}} \quad (10)$$

Based on relation (9), we can calculate the quenching constants in different concentration series solutions, which are listed in table 1, and it is found that k_{SV} decreases with the decrease of the acetic acid solution concentration.

Table1 k_{SV} of different concentration series

different concentration series	10^{-2}	10^{-3}	10^{-4}	10^{-5}	10^{-6}
k_{SV}	1.83	1.46	1.08	1.04	0.76

Based on the results shown in the reference [7], in some acetic acid solutions when fluorescence quenching occurs the ratio of $F_0/F = 2.81$, ($F_0/F - 1$) is equivalent to the quenching constants of k_{SV} the concentration series of 10^{-2} . As to the other concentration series the ratio of ($F_0/F - 1$) is also similar to the quenching constants k_{SV} . So we can conclude that the theoretical arithmetic of k_{SV} coincides with the experimental results.

3 Conclusions

From experiment results and above discussion, we can draw the conclusions:

Acetic acid solutions can absorb 253.7nm UV light and emit fluorescence with the peak wavelength at 395nm. Fluorescence intensities change with the solution concentrations, and fluorescent quenching occurs

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由表中数据可以看出,OE双输出棱镜分束角的测试值与理论计算值之差在6%之内,因此可以认为二者是一致的。

3.2 光强分束比的测试

OE双输出棱镜光强分束比的测试光路如图3所示。样品棱镜放在测角仪的中部,以调整光在棱

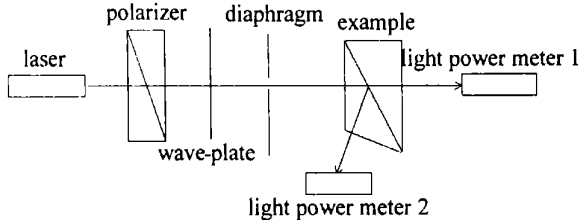


Fig. 3 Measurement setup of intensity splitting ratio

镜上的入射角。测角仪角度的读出精度为 $15'$;两功率计是同一型号并且经过中国计量科学研究院对633nm进行了定标。光经过起偏器与 $1/4$ 波片后成为圆偏振光(以消除He-Ne激光器出射光的部分偏振对测量的影响),经样品棱镜起偏分束后,由光功率计1和光功率计2同时探测出射的o,e光的光强 I_o 和 I_e ,然后由(10)式得出光强分束比的实验值如图4所示。

图4中曲线C表示实验值,曲线D表示理论计算值。由于测试条件的限制,没有在更多的光源中做测试验证。但由上图可知,对633nm光源而言,OE双输出棱镜光强分束比的实测值与理论计算值是基本一致的;对于OE双输出棱镜的光强分束比,正的入射角的影响要比负的入射角大,这是在使用这种棱镜时需要特别注意的。

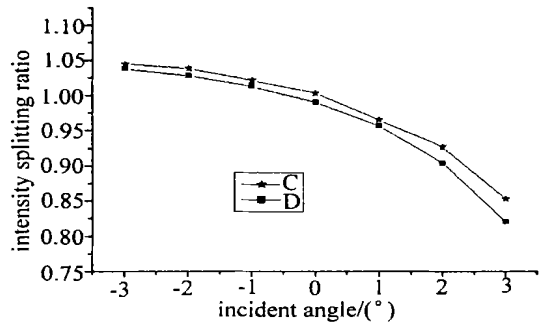


Fig. 4 The changeable relation of intensity splitting ratio with the incident angle

4 结论

就OE双输出棱镜的分束角而言,理论分析及实验测试的结果均表明:当入射角变化为 2° 时,分束角的变化为 $2'$,变化关系式为 $\theta = 180^\circ - 2S - 2\alpha$,其中 S 为棱镜的结构角。

OE双输出棱镜的光强分束比是随着入射角的改变呈现一种非线性的变化:当光正入射时($\alpha = 0$),e,o光之光强分束比(I_e/I_o)约为1;当 α 为负值时, $I_e/I_o > 1$;当 α 为正值时, $I_e/I_o < 1$ 。

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in some acetic acid solutions. The quenching mechanism is owing to singlet-triplet transition and k_{SV} of different concentration series is calculated respectively. Even in very diluent solutions, acetic acid can emit strong fluorescence. The reason is that acetic acid molecules form a kind of planar and rigid structure joined by hydrogen bond and this structure enhances fluorescent emission.

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